Protocols for Laboratory Verification of Performance of the BioFire® FilmArray® Blood Culture Identification (BCID) Panel

TECHNICAL
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A Laboratory Protocol for Use with Live Organisms

Purpose

The Clinical Laboratory Improvement Amendments (CLIA), passed in 1988, establishes quality standards for all laboratory testing to ensure the accuracy and reliability of patient test results, regardless of where the test is performed. The CLIA regulations include a requirement for verifying the performance specifications of unmodified, moderate complexity tests cleared or approved by the FDA.

This document provides an example of a verification procedure to assist your laboratory in developing a protocol for the verification of BioFire BCID Panel performance on BioFire® FilmArray® Systems as required by CLIA. A verification scheme, compatible with the BioFire BCID Panel, has been designed using non-clinical specimens. This scheme provides positive and negative tests for each organism detected by the BioFire BCID Panel and may be easily modified or expanded to meet specific criteria. Day-to-day variation is evaluated by testing each sample on two separate days. To evaluate user-to-user variation, multiple laboratory technicians may test the same sample. In addition, testing patient samples for verification of the performance of the BioFire BCID Panel should be done under the guidance of the Laboratory Director, but is not described here.

As per the CLIA regulation, the Laboratory Director is ultimately responsible for ensuring that verification procedures meet the appropriate standards for CLIA and applicable laboratory accrediting agencies.

FilmArray Intended Use

The BioFire BCID Panel is a qualitative multiplexed nucleic acid-based *in vitro* diagnostic test intended for use with BioFire Systems. The BioFire BCID Panel is capable of simultaneous detection and identification of multiple bacterial and yeast nucleic acids and select genetic determinants of antimicrobial resistance. The BioFire BCID Panel assay is performed directly on blood culture samples identified as positive by a continuous monitoring blood culture system. Results are intended to be interpreted in conjunction with Gram stain results.

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Technical Note
BioFire Diagnostics, LLC
www.biofiredx.com

QS-339B-01

The following gram-positive bacteria, gram-negative bacteria, and yeast are identified using the BioFire® FilmArray® Blood Culture Identification (BCID) Panel: Enterococci, *Listeria monocytogenes*, Staphylococci (including specific differentiation of *Staphylococcus aureus)*, Streptococci (with specific differentiation of *Streptococcus agalactiae*, *Streptococcus pneumoniae*, and *Streptococcus pyogenes*), *Acinetobacter baumannii*, *Enterobacteriaceae* (including specific differentiation of the *Enterobacter cloacae* complex, *Escherichia coli*, *Klebsiella oxytoca*, *Klebsiella pneumoniae*, *Proteus,* and *Serratia marcescens*), *Haemophilus influenzae*, *Neisseria meningitidis* (encapsulated), *Pseudomonas aeruginosa*, *Candida albicans*, *Candida glabrata*, *Candida krusei*, *Candida parapsilosis*, and *Candida tropicalis*.

The BioFire BCID Panel also contains assays for the detection of genetic determinants of resistance to methicillin (*mecA*), vancomycin (*vanA* and *vanB*), and carbapenems (bla_{KPC}) to aid in the identification of potentially antimicrobial resistant organisms in positive blood culture samples.

The complete intended use statement and additional information about the use of the BioFire® FilmArray® System can be found in the *BioFire® FilmArray® Blood Culture Identification (BCID) Panel Instruction Booklet*

Performance Verification: Overview

The procedure described below will generate multiple positive and negative results for each of the BioFire BCID Panel assays. The procedures were developed using organism strains available from Microbiologics®, Saint Cloud, MN (part numbers listed in materials section).

A simple procedure has been designed to take advantage of the multiplex nature of the BioFire BCID Panel. Verification testing efficiency is maximized by evaluating multiple target organisms in a single test run.

A BioFire System is defined as all BioFire® FilmArray® Instruments or modules that are connected to and controlled by a single computer system. If the laboratory director chooses not to perform the verification protocol on each individual instrument, it is advised that test replicates are evenly distributed among the instruments or modules.

Clinical/patient specimens may be used in place of or in addition to the verification schemes described here to assess clinical sensitivity and sample matrix effects in the performance verification of the BioFire BCID Panel.

Table 1. Overview of Verification Protocol

Verification Protocol	Organisms per Pool[a]	Number of Sample Pools	Replicates per Sample Pool	Pouches Required	Expected Positive Results	Expected Negative Results	Approximate Days of Testing[b]
Simple protocol[c]	6, 7, or 9	3	4	12	4 per organism	8 per organism	2

[a] Depending on the material used for verification, pooling of organisms may not be appropriate and the values in the table may need to be modified.
[b] The approximate number of days for testing assumes a system configured with one instrument/module and does not include time to grow microbial cultures.
[c] This simple protocol may be easily expanded to increase the number of pouches tested on one instrument or for the verification of multiple instruments.

Performance Verification: Materials

The following materials may be used to perform verification procedures:

Table 2. Materials needed for recommended verification procedure

Material	Part Number
BioFire® FilmArray® Blood Culture Identification (BCID) Panel (30 tests per kit)	BioFire Diagnostics, LLC RFIT-ASY-0126
BioFire® FilmArray® Blood Culture Identification (BCID) Panel Instruction Booklet	BioFire Diagnostics, LLC (RFIT-PRT-0369)
BioFire® FilmArray® Blood Culture Identification Panel Quick Guide	BioFire Diagnostics, LLC (RFIT-PRT-0370)
Blood culture media[a]	BD BACTEC™ Plus Aerobic/F Medium (with resin) BD, 442192 (or equivalent)
Human Whole Blood with EDTA (pathogen free)	Bioreclamation LLC, HMWBEDTA2 (or equivalent, with anticoagulant)
McFarland Turbidity Standard, 1.0	Fisher Scientific, R20411 (or equivalent)
Phosphate Buffered Saline, pH 7.4	Sigma, P3813 (or equivalent)
Polystyrene tube with cap (12 mL, 16 x 100 mm, round-bottom)	VWR, 82050-246 (or equivalent)
Polypropylene centrifuge tube with flat cap (50 mL, sterile)	VWR, 89004-364 (or equivalent)

[a] See Table 77 in the BioFire® FilmArray® Blood Culture Identification (BCID) Panel Instruction Booklet for other acceptable blood culture media/bottle types.

Table 3. Recommended organism strain and source for verification procedure

Organism	Microbiologics Catalog Number[a]
Acinetobacter baumannii ATCC® 19606™ KWIK-STIK	0357P
Candida albicans ATCC® 10231™ Lab-Elite	0443-CRM
Candida glabrata ATCC® 15126™ KWIK-STIK	0737P
Candida krusei ATCC® 14243™ KWIK-STIK	0809P
Candida parapsilosis ATCC® 22019™ KWIK-STIK	0726P
Candida tropicalis ATCC® 1369™ KWIK-STIK	01036P
Enterobacter cloacae subsp. cloacae ATCC® 13047™ Lab-Elite	0323-CRM
Enterococcus faecalis ATCC® 51299™ KWIK-STIK	0959P[b]
Escherichia coli ATCC® 11229™ Lab-Elite	0681-CRM
Haemophilus influenzae ATCC® 10211™ KWIK-STIK	0441P
Klebsiella oxytoca ATCC® 13182™ KWIK-STIK	0530P
Klebsiella pneumoniae ATCC® BAA-1705™ KWIK-STIK	01005P[c]
Listeria monocytogenes ATCC® 19111™ KWIK-STIK	0277P
Neisseria meningitidis ATCC® 13077™ KWIK-STIK	0453P
Proteus mirabilis ATCC® 35659™ Lab-Elite	0944-CRM
Pseudomonas aeruginosa ATCC® 27853™ KWIK-STIK	0353P
Serratia marcescens ATCC® 13880™ KWIK-STIK	0247P
Staphylococcus aureus subsp. *aureus* ATCC® 33591™ Lab-Elite	0496-CRM[d]
Staphylococcus epidermidis ATCC® 12228™ Lab-Elite	0371-CRM
Streptococcus agalactiae ATCC® 12386™ KWIK-STIK	0439P
Streptococcus pneumoniae ATCC® 10015™ KWIK-STIK	0865P
Streptococcus pyogenes ATCC® 19615™ Lab-Elite	0385-CRM
BioFire® FilmArray® Blood Culture Identification (BCID) Verification Panel	5229P[e]

[a] Any appropriate source of organism may be used for verification of any or all of the assays in the BioFire BCID Panel. However, when alternate organism sources are used, the sample volumes or pooling schemes suggested in the examples below may need to be adjusted. Alternate organism strains may not provide the same results for antimicrobial resistance genes as those suggested here.

[b] This strain of *E. faecalis* (ATCC® 51299) carries the *vanB* gene (vancomycin resistance) .

[c] This strain of *K. pneumoniae* (ATCC® BAA-1705) carries the bla_{KPC} gene (carbapenems resistance).

[d] This strain of *S. aureus* subsp. *aureus* (ATCC® 33591) carries the *mecA* gene (methicillin resistance).

[e] This is a bundled part number containing all the organisms listed above.

TECHNICAL ::: NOTE

Performance Verification Protocol

The protocol described below utilizes samples prepared by pooling together up to 9 different organism suspensions in a simulated blood culture matrix. The pooling scheme (Table 4) should be followed to obtain the expected number of positive and negative results for each organism in a time and resource-efficient manner.

Note: Dilution of organisms beyond levels proposed in these guidelines may lead to inconsistent results and is not recommended.

The protocol can be followed to test a total of 12 pooled samples, providing 4 positive results and 8 negative results per organism. This example demonstrates how the verification can be completed by performing six tests per day. This testing scheme can be modified to run more samples per day based on the number of instruments in the BioFire® FilmArray® System. The number of samples tested per day should be determined by the individual laboratory.

Table 4. Recommended Organism Pooling Scheme

Organism	Organism Volume	Human Whole Blood	BD Culture Medium	Approximate Final Volume of Pool
Pool 1				
Candida albicans	0.1 mL	3 mL	8 mL	~ 12 mL
Candida krusei	0.1 mL			
Streptococcus agalactiae	0.1 mL			
Neisseria meningitidis	0.1 mL			
Pseudomonas aeruginosa	0.1 mL			
Staphylococcus aureus (MRSA)*	0.1 mL			
Streptococcus pyogenes	0.1 mL			
Pool 2				
Enterococcus faecalis	0.2 mL	3 mL	8 mL	~ 12 mL
Staphylococcus epidermidis (MSSE)**	0.4 mL			
Acinetobacter baumannii	0.1 mL			
Candida glabrata	0.1 mL			
Candida tropicalis	0.2 mL			
Enterobacter cloacae	0.3 mL			
Klebsiella oxytoca	0.1 mL			
Listeria monocytogenes	0.1 mL			
Escherichia coli	0.1 mL			
Pool 3				
Candida parapsilosis	0.1 mL	3 mL	8 mL	~ 12 mL
Klebsiella pneumoniae	0.1 mL			
Proteus mirabilis	0.1 mL			
Serratia marcescens	0.1 mL			
Haemophilus influenzae	0.1 mL			
Streptococcus pneumoniae	0.1 mL			

*MRSA, methicillin resistant *S. aureus*.
**MSSE, methicillin susceptible *S. epidermidis*.

Protocol Example

The estimated total time to completion for this verification example is 2 days for systems configured with one instrument (not including time to grow microbial cultures). A proposed organism pooling scheme is presented above in Table 4.

TECHNICAL
::: NOTE

Day 1

1. Obtain a pure culture of each organism that has been streaked for isolation on agar media appropriate for the organism. It is recommended to use agar plate cultures that are less than 1 week old. See Microbiologics product insert for use of KWIK-STIK cultures.

2. Prepare a suspension of each organism equivalent to McFarland turbidity standard 1.0 using approximately 3 mL of phosphate buffered saline (PBS), pH 7.4.

3. Prepare three sample pools according to the organism pooling scheme presented in Table 4. The sample pool preparation worksheet in the Appendix can assist in the set-up to ensure all components are added to each pool.

   Note: It is important to prepare only the number of sample pools that will be tested within 2 days. The suggestion to prepare 3 sample pools is based on testing 6 pouches per day using one instrument. The number of samples prepared may be increased or decreased based on the laboratory's work schedule and the number of instruments connected within a BioFire® FilmArray® Instrument.

   a. Use a pipette to add 3 mL of human whole blood to a 50 mL conical tube.

   Note: It is recommended to use blood that has been prescreened as negative for BioFire® FilmArray® Blood Culture Identification (BCID) Panel pathogens.

   b. Use a 12 gauge needle and a syringe to remove 8 mL of blood culture medium from a blood culture bottle and add it to the conical tube containing whole blood. Care should be taken to minimize transferring resin beads into the sample.
   c. Use a pipette to transfer the appropriate volume (Table 4) of organism suspension to the blood-medium mixture.
   d. Repeat for the remaining organisms in the pool to combine the appropriate organisms into a single 50 mL tube.
   e. Ensure the pooled sample is effectively mixed by vortexing prior to removing a sample for testing.
   f. Refrigerate samples (2–8°C) for up to 2 days for the evaluation of day-to-day variation.

4. Prepare and test two samples (i.e. A and B, see Figure 1) from a single sample pool (i.e. Pool #1). The duplicate samples should be tested in a single day by different users.

   Note: For each sample, follow instructions in the *BioFire® FilmArray® Blood Culture Identification (BCID) Panel Instruction Booklet* and *BioFire® FilmArray® Blood Culture Identification (BCID) Panel Quick Guide* for pouch preparation, pouch hydration, sample loading, and sample testing.

5. Repeat Step 3 for the remaining sample pools (pools #2 and #3) to be tested that day.

Day 2

To evaluate day-to-day variation, test the remaining samples (i.e. samples C and D) from the same sample pools prepared on Day 1 by repeating Step 3 above.

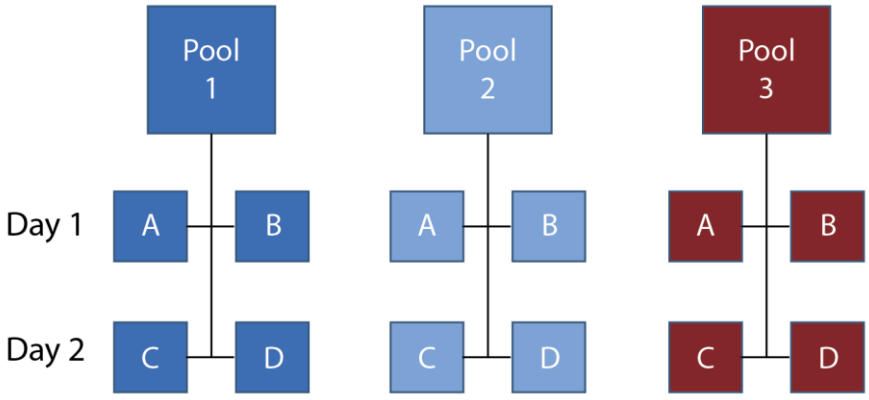

Figure 1. Workflow for example protocol

Expanding the protocols

The protocol described above can be expanded by increasing the number of tests from each of the organism pools. Each organism pool contains approximately 12 mL, which is enough material to complete many tests for each pool.

Verification of Loaner, Repaired, and Permanent Replacement Instruments

If it becomes necessary to verify the performance of a loaner, repaired, or permanent replacement instrument, the following protocol may serve as a guideline but should be verified by the Laboratory Director.

1. Select a few specimens and/or proficiency samples (any combination of positives and negatives) previously tested on the BioFire® FilmArray® Blood Culture Identification (BCID) Panel. The Laboratory Director should determine the appropriate number of samples to test. Proficiency samples should not be pooled or diluted.
2. Test the selected specimens/samples on the loaner or repaired instrument and document the results.

Technical Support Contact Information

BioFire is dedicated to providing the best customer support available. If you have any questions or concerns about this process, please contact the BioFire Technical Support team for assistance.

BioFire Technical Support
Email: support@biofiredx.com
Phone: +1-801-736-6354, select Option 5

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Appendix

Sample Pool Preparation Worksheet

Organism	Organism Volume	Human Whole Blood Volume	Blood Culture Medium Volume	Approximate Final Volume of Pool
Pool 1				
Candida albicans	☐ 0.1 mL	☐ 3 mL	☐ 8 mL	~ 12 mL
Candida krusei	☐ 0.1 mL			
Streptococcus agalactiae	☐ 0.1 mL			
Neisseria meningitidis	☐ 0.1 mL			
Pseudomonas aeruginosa	☐ 0.1 mL			
Staphylococcus aureus (MRSA)*	☐ 0.1 mL			
Streptococcus pyogenes	☐ 0.1 mL			
Pool 2				
Enterococcus faecalis	☐ 0.2 mL	☐ 3 mL	☐ 8 mL	~ 12 mL
Staphylococcus epidermidis (MSSE)**	☐ 0.4 mL			
Acinetobacter baumannii	☐ 0.1 mL			
Candida glabrata	☐ 0.1 mL			
Candida tropicalis	☐ 0.2 mL			
Enterobacter cloacae	☐ 0.3 mL			
Klebsiella oxytoca	☐ 0.1 mL			
Listeria monocytogenes	☐ 0.1 mL			
Escherichia coli	☐ 0.1 mL			
Pool 3				
Candida parapsilosis	☐ 0.1 mL	☐ 3 mL	☐ 8 mL	~ 12 mL
Klebsiella pneumoniae	☐ 0.1 mL			
Proteus mirabilis	☐ 0.1 mL			
Serratia marcescens	☐ 0.1 mL			
Haemophilus influenzae	☐ 0.1 mL			
Streptococcus pneumoniae	☐ 0.1 mL			

*MRSA, methicillin resistant *S. aureus*.
**MSSE, methicillin susceptible *S. epidermidis*.

BioFire® FilmArray® Instrument Verification Record

Computer System Serial # _____

BioFire® FilmArray® Blood Culture Identification (BCID) Panel, Kit Part #: _____ Lot #: _____

Organism/Sample Source and Lot #: _____

TECHNICAL ::: NOTE

Organism	Instrument Serial #	Was the Organism Detected?	No. Positive	No. Negative	No. Days Tested	No. Users	Patient Samples Tested?
Acinetobacter baumannii		☐ Yes ☐ No					
Candida albicans		☐ Yes ☐ No					
Candida glabrata		☐ Yes ☐ No					
Candida krusei		☐ Yes ☐ No					
Candida parapsilosis		☐ Yes ☐ No					
Candida tropicalis		☐ Yes ☐ No					
Enterobacter cloacae		☐ Yes ☐ No					
Enterococcus faecalis (with vanA/B call)		☐ Yes ☐ No					
Escherichia coli		☐ Yes ☐ No					
Haemophilus influenzae		☐ Yes ☐ No					
Klebsiella oxytoca		☐ Yes ☐ No					
Klebsiella pneumoniae (with KPC call)		☐ Yes ☐ No					
Listeria monocytogenes		☐ Yes ☐ No					
Neisseria meningitidis		☐ Yes ☐ No					
Proteus mirabilis		☐ Yes ☐ No					
Pseudomonas aeruginosa		☐ Yes ☐ No					
Serratia marcescens		☐ Yes ☐ No					
Staphylococcus aureus (MRSA, with mecA call)		☐ Yes ☐ No					
Staphylococcus epidermidis (MSSE, no mecA call)		☐ Yes ☐ No					
Streptococcus agalactiae		☐ Yes ☐ No					
Streptococcus pneumoniae		☐ Yes ☐ No					
Streptococcus pyogenes		☐ Yes ☐ No					

Reviewed by: _____

Signature Date